

Product Sheet

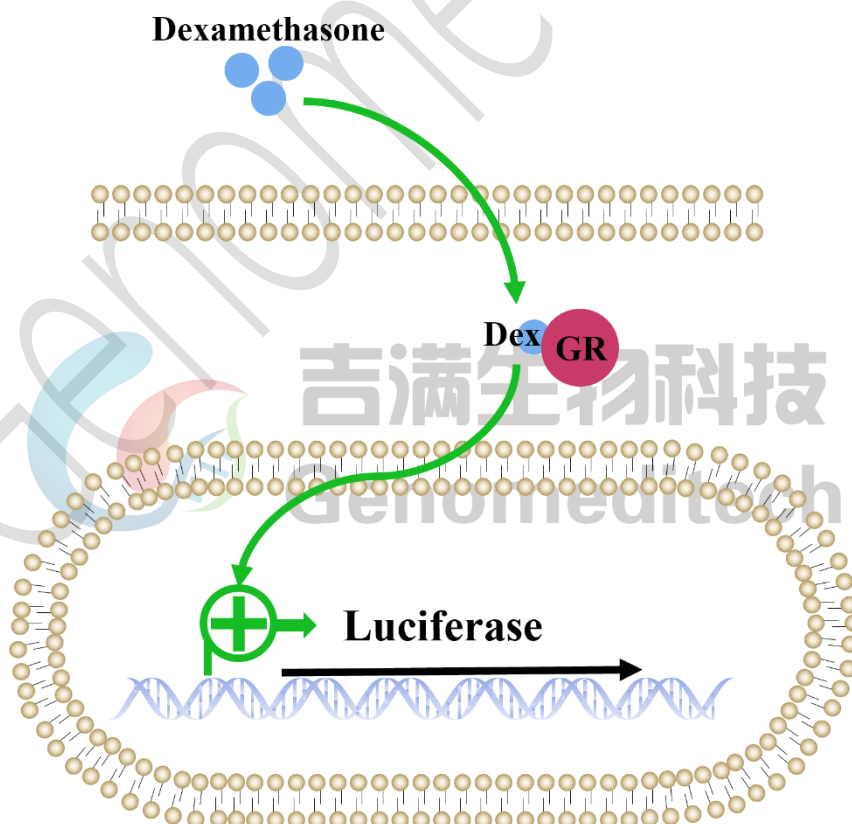
GRE-Luc Reporter Hela Cell Line

Catalog number: GM-C40521

Version 3.3.1.251017

The glucocorticoid receptor (GR) is a ligand-dependent transcription factor belonging to the nuclear receptor superfamily, and it is widely expressed in various human tissues. GR primarily mediates the biological effects of glucocorticoids, such as the regulation of metabolism, immune responses, inflammation, and stress reactions. Upon binding to glucocorticoids, GR undergoes a conformational change and dissociates from its chaperone proteins (such as HSP90), after which it translocates into the nucleus. Once inside the nucleus, GR can directly bind to glucocorticoid response elements (GREs) to regulate the transcription of target genes, or indirectly modulate gene expression through interactions with other transcription factors (such as NF- κ B and AP-1).

GRE-Luc Reporter Hela Cell Line is a clonal stable Hela cell line constructed using lentiviral technology, endogenously expresses Glucocorticoid Receptor gene, along with signal-dependent expression of a luciferase reporter gene. After stimulation with dexamethasone, it activates downstream signaling pathways, leading to the expression of luciferase. The luciferase activity measurement indicates the activation level of the signaling pathway and therefore be used to evaluate the in vitro effects of related drugs.



Specifications

Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt

Recovery Medium	DMEM/F12+10% FBS+1% P.S
Growth medium	DMEM/F12+10% FBS+1% P.S+0.5 µg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO ₂

Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
DMEM/F12	Gibco/C11330500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/ GM-040401
Dexamethasone	Beyotime/ST1258
GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit	Genomeditech/ GM-040513

Figures

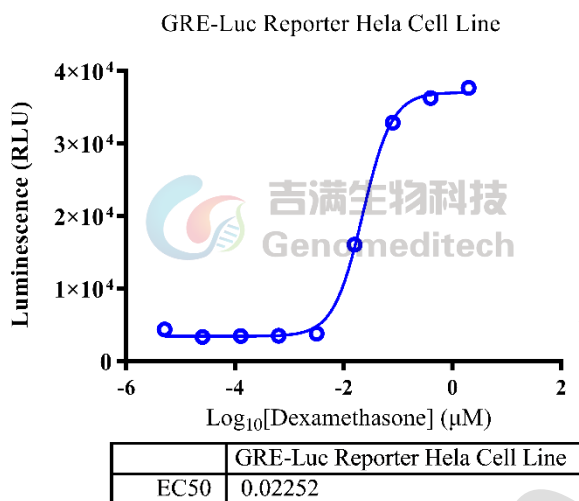


Figure 1 | Response to Dexamethasone. The GRE-Luc Reporter Hela Cell Line (Cat. GM-C40521) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Dexamethasone (Beyotime/ST1258) in assay buffer (DMEM/F12+1% FBS+1% P.S) for 16 hours. The firefly luciferase activity was measured using the Luciferase Reporter Assay Kit (Genomeditech). The maximum induction fold was approximately [9.6]. Data are shown by drug molar concentration.

Cell Recovery

Recovery Medium: DMEM/F12+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at 176 x g for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: DMEM/F12+10% FBS+1% P.S+0.5 µg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:2 to 1:3 every 2 to 3 days.
- b) Remove and discard culture medium.
- c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes at 37°C).
- e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 - 1:3 is recommended

Medium Renewal: Every 2 to 3 days

Notes

- a) Fetal bovine serum (FBS) needs to be heat-inactivated at 56°C for 30 minutes, which can inactivate complement and certain viruses without significantly affecting the activity of most growth factors and cytokines.

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